

The quest for a prophylactic HIV vaccine continues: results from a phase I trial using novel routes of DNA vaccination in HIV uninfected volunteers

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Background

- As the HIV epidemic continues to expand, it is widely accepted a preventative HIV vaccine is a vital public health priority
- DNA vaccines are being increasingly utilized based on several advantages over other vaccines including rapid manufacture, cost effectiveness and lack of concerns associated with the administration of infectious agents¹
- Limitations to DNA vaccination are related to inefficient uptake of DNA into cells and rapid degradation, thus ineffective antigen presentation
- Ongoing efforts investigating DNA vaccine delivery are therefore of great interest in an effort to improve immunogenicity and these include cutaneous routes
- Previous studies have shown a dose sparing effect of intradermal (ID) vaccination, where only 10-20% of a conventional intramuscular dose (IM) is needed to be adequately immunogenic²
- Transcutaneous (TC) vaccine delivery, is 'needle-free' and in a recent Phase I clinical trial using an influenza/tetanus vaccine³, this method favoured the development of CD8+ T cell responses, a potentially crucial killing mechanism in HIV. This method has never been utilised in a DNA preventative HIV vaccine trial
- Physical methods of delivering DNA such as in vivo electroporation (EP) have been shown to enhance DNA immunogenicity in early clinical trials⁴
- In this Phase I RCT we report on the safety and immunogenicity of a GTU[®] MultiHIV B clade DNA vaccine in HIV uninfected participants administered via the 3 different routes described above

Methods

- 30 HIV uninfected participants deemed eligible according to strict criteria, were randomised to received GTU[®] MultiHIV Clade B DNA vaccine at weeks 0/4/12 by 1 of 3 routes: IM+ID, IM+TC and IM+EP
- Participants were assessed for local and systemic reactogenicity and adverse events
- T cell responses to vaccine encoded peptides (*rev*, *tat*, *nef*, *gag*, *CTL* coded for by *pol* and *env* genes) made were measured by IFN- γ ELISpot and intracellular cytokine staining (ICS)
- An exploratory end point to assess CD8+ T cell function was conducted using a viral inhibition assay (VIA)

References:
 1. Martinon et al. Persistent immune responses induced by a human immunodeficiency virus DAN vaccine delivered in association with electroporation in the skin of non human primates. *Hum Gen Ther* 2009;20:1291-1307
 2. Kenney RT, Frech SA, Muenz LR, Villar CP, Glenn GM. Dose sparing with intradermal injection of influenza vaccine. *N Engl J Med* 2004;351:2295-2301.
 3. Combadiere B, Vogt A, Mahe B, Costagliola D, Hadam S, Bonduelle O, et al. Preferential amplification of CD8 effector-T cells after transcutaneous application of an inactivated influenza vaccine: a randomized phase I trial. *PLoS One* 2010;5:e10818.
 4. Vasan S, Hurley A, Schlesinger SJ, Hannaman D, Gardiner DF, Dugin DP, et al. In vivo electroporation enhances the immunogenicity of an HIV-1 DNA vaccine candidate in healthy volunteers. *PLoS One* 2011;6:e19252.
 5. Spentzou A, Bergin P, Gill D, Cheeseman H, Ashraf A, Kaltsidis H, et al. Viral inhibition assay: a CD8 T cell neutralization assay for use in clinical trials of HIV-1 vaccine candidates. *J Infect Dis* 2010;201:720-729.

Results

Recruitment and participant flow

59 participants were screened with 30 deemed eligible according to inclusion/exclusion criteria. The table below shows the demographics of enrolled participants (Table 1)

Demographics	Group 1 ID+IM	Group 2 TC+IM	Group 3 EP+IM	Total (%)
Gender				
Male	5	6	6	17 (56)
Female	4	5	4	13 (43)
Ethnicity				
White	8	7	8	23 (77)
Black	0	2	2	4 (13)
Asian	0	2	0	2 (7)
Other Ethnic Group	1	0	0	1 (3)
Age (years)				
Mean (range)	27.4 (22-34)	31.5 (21-42)	31.7 (23-39)	30.4 (21-42yrs)

Reactogenicity and safety

- Overall there were no safety concerns with no significant differences between groups with respect to local and systemic reactogenicity (Fig 1A). In the EP+IM group there were a greater number of participants documenting pain in the leg on their diary cards following EP (Fig 1B), although over 90% participants in this group thought it to be an acceptable route of vaccine delivery.
- In the TC+IM group, a greater number of participants documented redness and discoloration secondary to the procedure compared to the ID+IM group (7 responses compared to 2 respectively Fig 1C). 5/11 participants in the TC group had either hypo or hyperpigmentation related to post-inflammatory changes as a direct result of the TC procedure (example shown in Fig 2). All these changes resolved over time, although notably 1 person in the TC+IM group declined the final vaccination as a result of ongoing hypopigmentation over the vaccine site

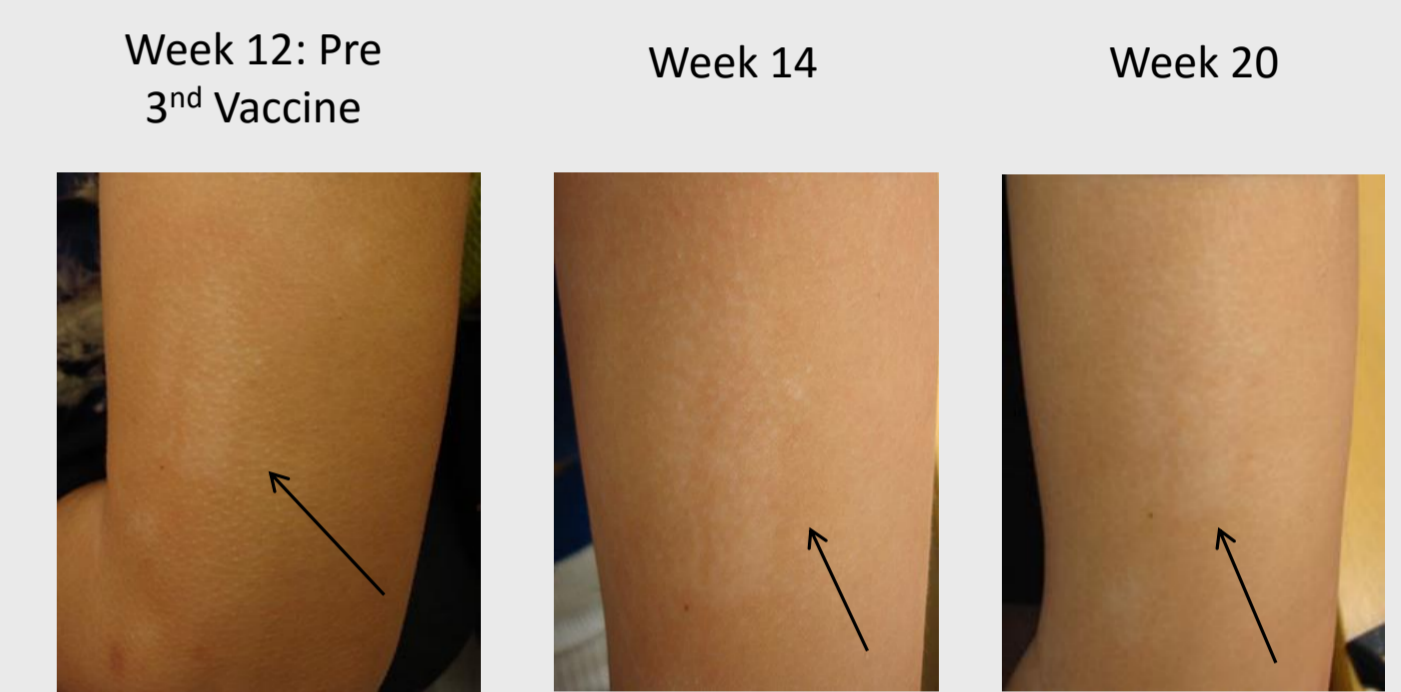
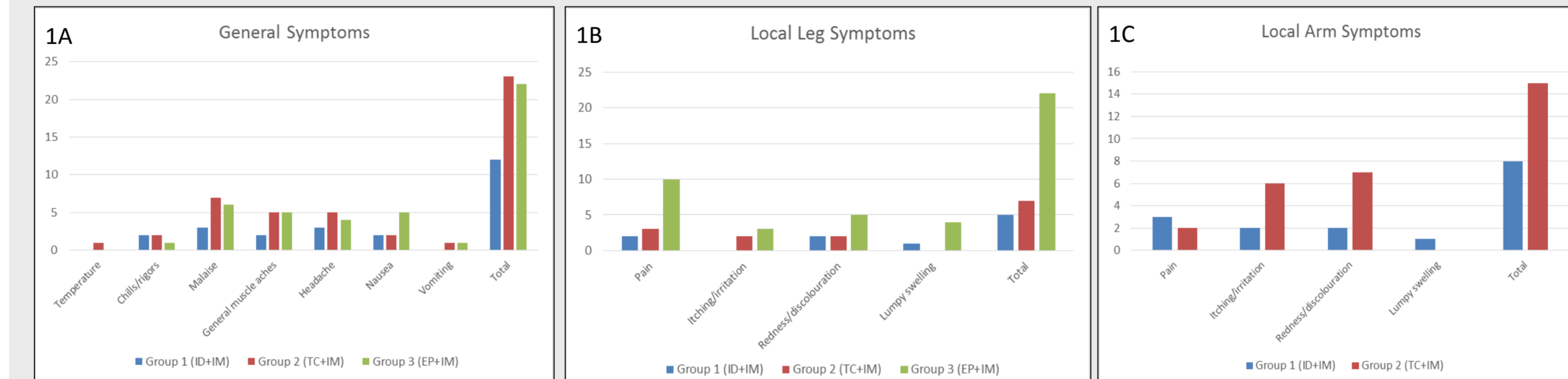


Figure 2: Example of post inflammatory hypopigmentation as a result of TC procedure. This participant was Caucasian and declined the final vaccine due to persisting changes.

Immunogenicity

1. T Cell ELISpots (primary end point = 2 weeks post final vaccine)

- At the primary end point, EP+IM performed better than both ID+IM and TC+IM groups with 9/10 responders to any peptide in the EP+IM group compared to 1/9 in the ID+IM group and 0/11 in the TC+IM group (Fig 3A). Pooled peptide responses were also greater in the EP +IM group (Fig 3B)
- Within the EP+IM group, the greatest magnitude of IFN- γ response was to Nef and Gag peptide pools with 8/10 (80%) and 9/10 (90%) participants responding to these antigens respectively (Figure 3C). The weakest responses were to Tat and this was consistent across all 3 vaccination groups. The EP+IM group also showed duration of response across time (Figure 3D).

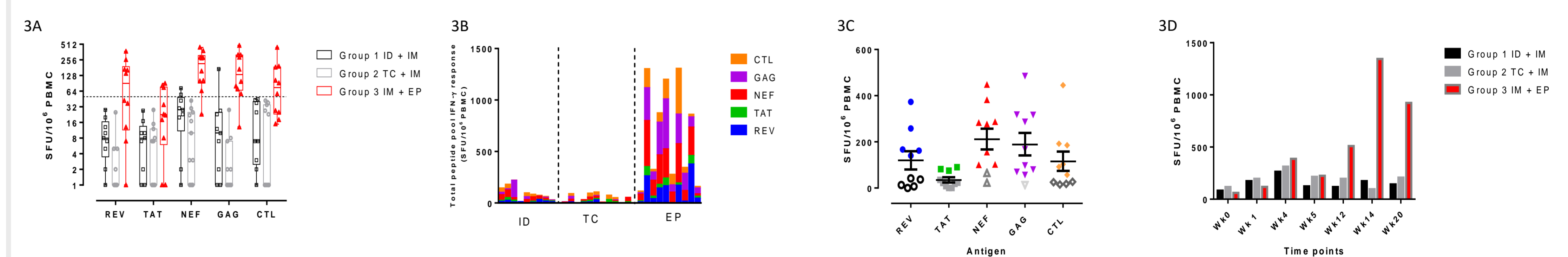


Figure legend: 3A-Primary end point data comparing IFN- γ response to all peptide pools. Dotted line denotes cut off for positive response (>55 SFU/M PBMC). 3B-pooled peptide response by group at the primary end point. 3C- EP+IM group only showing IFN- γ response to the peptides. 3D- IFN- γ response in different groups across time.

2. Intracellular cytokine staining (ICS)

- ICS analysis showed a significant increase in CD4+ gag specific IFN- γ responses in the IM+EP group, with few CD8+ specific responses (Fig 4)

3. Viral inhibition assay (VIA)

- A viral inhibition assay was used to measure the in vitro ability of CD8+ cells to inhibit HIV replication based on a previously published assay⁵ and using a panel of 6 viruses (Table 2)
- All groups showed viral inhibitory activity to at least one virus at the primary end point, in 2/9, 4/9 and 5/7 participants across groups ID+IM, TC+IM, EP+IM respectively.
- The greatest number of participants with detectable HIV-specific CD8+ cells capable of inhibiting any virus was in the EP+IM group (71% participants) and to the greatest number of viruses in the panel (4/6), with the greatest cross clade inhibition (Figs 5A and 5B).

Figure legend: 4A- ICS data showing CD4+ IFN- γ response at primary end point 4B- ICS data showing CD8+ IFN- γ response at primary end point. 5A- Viral inhibition assay showing inhibition in red according to IAVI criteria (>1.5 log and >0.6 baseline inhibition). 5B- No. participants inhibiting virus by group showing clade B and non clade B viruses

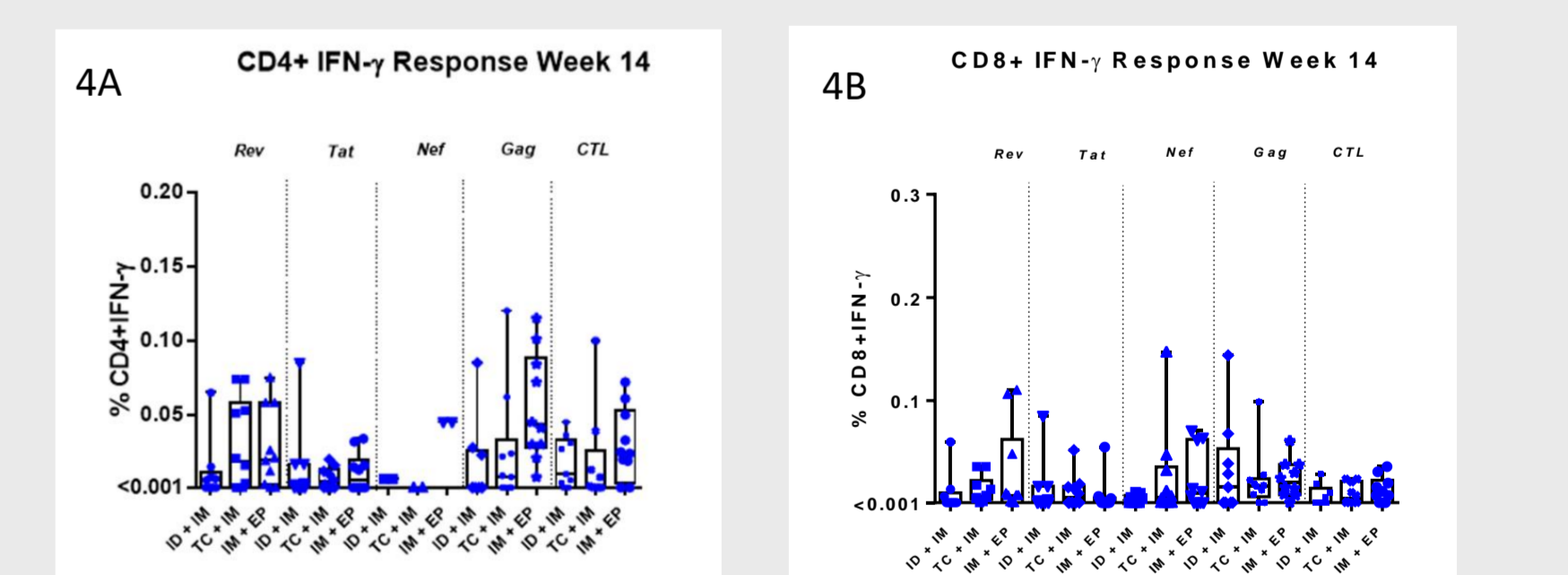
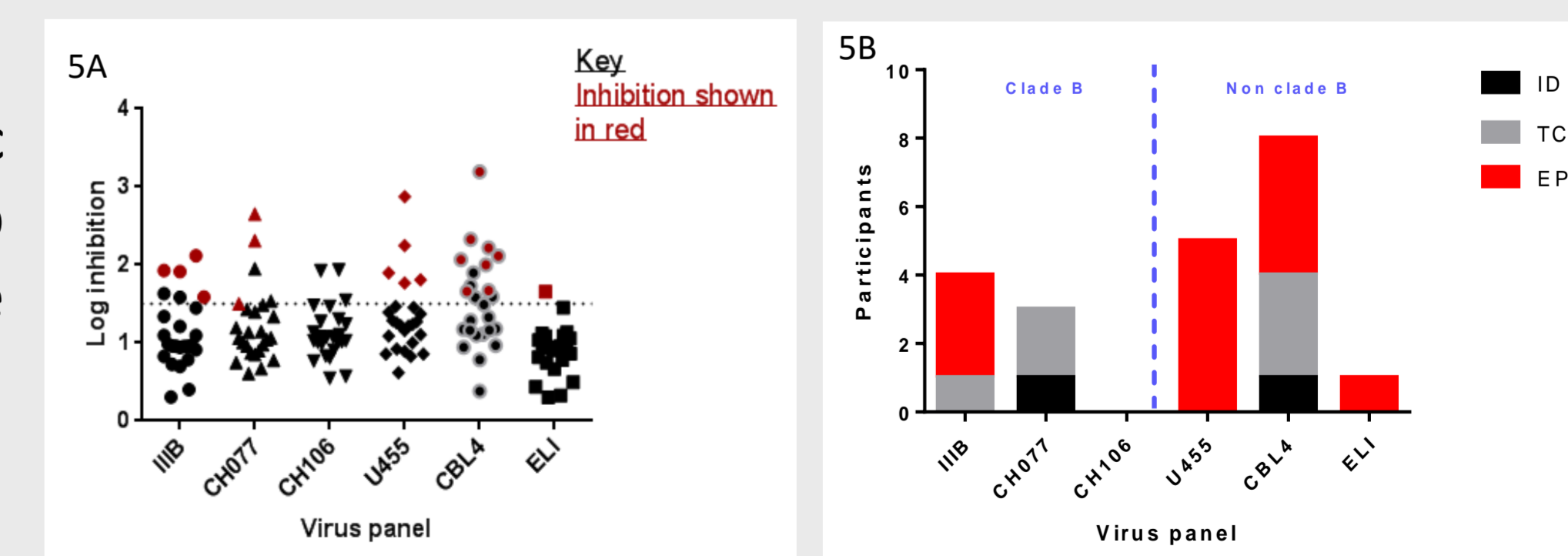


Table 2: Panel of 6 viruses using in VIA

VIRUS	CLADE	TYPE	CKCR4/CCRS
11IB	B	Lab	X4
CH077	B	IMC	R5
CH106	B	IMC	R5
ELI	AD	Lab	X4
U455	A	Lab	X4
CBL4	D	Lab	X4



Conclusions

- The GTU[®] MultiHIV B clade DNA vaccine was safe and well tolerated across all routes of vaccine administration and showed the greatest cellular immunogenicity when administered IM with EP
- The use of EP induced the greatest and broadest CD8+ viral inhibitory activity, although all groups also showed inhibition
- Importantly and in line with other clinical trials using EP, participants found the procedure to be tolerable and acceptable
- It is important to place this predominantly T cell based DNA vaccine in the context of wider clinical trials assessing combined T can B cells responses, in the ultimate goal of developing a robust HIV preventative vaccine